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Synthesis of Silver Nanoparticles by ecofriendly nvironmental method using Piper nigrum, Ziziphus spina-christi, and Eucalyptusglobulus extract

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Abstract. In the present study, silver nanoparticles (AgNPs) were prepared using an eco-friendly method synthesized in a single step biosynthetic using leaves aqueous extract of Piper nigrum , Ziziphus spina-christi, and Eucalyptus globulus act as a reducing and capping agents, as a function of volume ratio of aqueous extract(100ppm) to AgNO₃ (0.001M), (1: 10 ,2: 10 ,3: 10). The nanoparticles were characterized using UV-Visible spectra, X-ray diffraction (XRD). The prepared AgNPs showed surface Plasmon resonance centered at 443, 440,and 441 nm for sample prepared using extract Piper nigrum , Ziziphus spina-christi, and Eucalyptus respectively. The XRD pattern showed that the strong intense peaks indicate crystalline nature and face centered cubic structure of silver nanoparticles for all samples were prepared .The average crystallite size of the AgNPs was 20-45 nm. Morphology of the AgNPs were carried out using FESEM. Observations show that the AgNPs synthesized were spherical(Cluster) in shape. with diameters of 13 to 53 nm.

Key words ;AgNPs; Surface plasmon resonance; Piper nigrum ;Ziziphus spina-christi; Eucalyptus.

1. Introduction

Nanotechnology is considered to be one of the most important modern techniques of science. New properties of nanoparticles or nanomaterials such as morphology, particle size and distribution have enabled these materials to be an infrastructure in many applications, whether medical, such as those involved in drug delivery or biomedicine in the treatment of certain diseases and cosmetics in addition to environmental applications, and chemical industries, Catalysts, electronics, optics and other applications [1–3]. Silver nanoparticles (AgNPs) are unique nanoparticles with good chemical



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stability, unique antibacterial effect and anti-inflammatory activity [4, 5]. It can be used in many biomedical applications such as wound dressings, topical creams, disinfectant sprays and fabrics due to its antiseptic and biocide effect against microorganisms. They work by disrupting the unicellular membrane of microorganisms that thus disturb their enzymatic activities. In addition, recently, the synthesis of AgNPs for the diagnosis and treatment of cancer has received the attention of many scientists [6,7].

However, the synthesis of nanoparticles by chemical or physical methods is either expensive or associated with environmental risks due to the inclusion of certain toxic chemicals during the synthesis process. Biological Synthesis Inspired by Nanoparticles an alternative environmentally friendly process promised to synthesize nanoparticles because they do not involve the use of toxic chemicals during synthesis. Biological synthesis of nanoparticles is an environmentally friendly method involving the use of microorganisms [8-10], enzyme [11], and plant or plant extract [12]. Of all biological methods, the advantages of aggregate nanoparticles that use plants are to eliminate the complex process of preserving cell cultures [12]. In addition, it is a process capable of synthesizing nanoparticles. However, in order to replace chemical methods, biological methods must be developed in terms of accelerating the synthesis rate. On the other hand, due to serious issues, ad hoc research on the development of green chemistry and other biological processes has led to an environmentally friendly approach to the synthesis of mineral nanoparticles [13]. Plant sources such as leaves, bark, fruit extracts [14, 15] and organisms [16, 17] are commonly used in this approach. The plant extraction method is a cost-effective, time-efficient green method and provides a combination of crystalline nanoparticles of a wide range of sizes and shapes. Here, we used Papper, Rhamnus, and Eucalyptus act as a reducing and capping agents, and compared the composition of silver nanoparticles as a function of mole ratio aqueous extract to silver nitrate (AgNO_3) concentration (1: 10, 2: 10, 3: 10).

2. Experimental details

2.1 Materials and methods

2.1.1 Plant material Leaves of *Piper nigrum*, *Ziziphus spina-christi*, and *Eucalyptus globulus*. were obtained from local markets in Ramadi/ Iraq. Preparation of plant leaf extract 10 g powder of dry leaves were taken in a round flask along with 100 mL of Distilled water, allowed to stirrer at room temperature for 30 min under reflux condition. The extract obtained was filtered through Whatman No. 1 filter paper. The filtrate are drying and stored at 4 °C for further experiments.

2.1.2 Green synthesis of silver nanoparticles: For synthesis of silver nanoparticles, 10 ml of 1 mM AgNO_3 (Shanghai Jiuling Chemical Co., Ltd.) aqueous solution, were added to 1 ml (100ppm) of plant extract and left at room temperature for one hour. The change in color was observed indicating the formation of silver nanoparticles.

2.2 Characterization of green synthesis silver nanoparticles

Silver nanoparticle was characterized using an UV-vis spectrum for the detection of surface Plasmon resonance property (SPR) of AgNPs conducted at room temperature using Shimadzu UV-Vis 1800 spectrophotometer at wavelengths ranging from 300 to 800 nm using double beam UV-visible spectrophotometer (PD-303 UV).

The structure evolution of the green synthesized AgNPs using thymus vulgaris aqueous extract was examined by high-resolution X-ray diffraction (HR-XRD) using X'Pert Pro MRD diffractometer (PANalytical Company) system equipped with Cu-K α -radiation wavelength ($\lambda = 0.15418$ nm) operating at 40 kV and 30. Morphology and microstructure of the thin films were

investigated by scanning electron microscopy (FESEM) using Jeol JSM-6460 LV microscope operating at 10 kV.

3. Results and Discussions

3.1 Characterization of green synthesis gold nanoparticles.

3.1.1 UV spectral analysis

During the mixing process of the silver nitrate solution (colorless) and the pepper extract (greenish solution), the solution color changes due to the reaction process to yellowish brown. The change in color gives an initial indication of the formation of nanoparticle particles [9]. The appearance of silver particles in non-silver color is attributed to the Surface plasmon resonance. Surface plasmon resonance occurs in certain metals such as silver due to the arrival of its particle diameter to the nanometer. Therefore, the spectral analysis device is used at wavelengths of visible light to prove the formation of nanoparticles. The UV-Visible absorption spectra of the colloidal silver nanoparticles as a function of concentration of extract shown in Figure 1. An increase in absorbance can be noticed in the UV-Vis spectrum for AgNPs using *Piper nigrum* extract (Fig. 1.a). Also, the peak area at 439 to 443 nm has increased with increasing concentration of extract. The sharp peak at around 443 nm evidences the formation of silver nanoparticles [20], while the increase in intensity could be due to increasing number of nanoparticles formed as a result of reduction of silver ions present in the aqueous solution [21]. Figure 1.b show the UV-Vis spectrum for AgNPs prepared using *Ziziphus spina-christi*. It was observed that the peak area at 423 to 441 nm has increased with increasing concentration of extract, the peak intensity were less than that for simple prepared using *Ziziphus spina-christi* extract. The absorbance for AgNPs were prepared using extract, *Eucalyptus globulus* show increase intensity compared with the other sample (Fig1.c). It noticed that the peak area at 432 to 440 nm has increased with increasing concentration of extract.

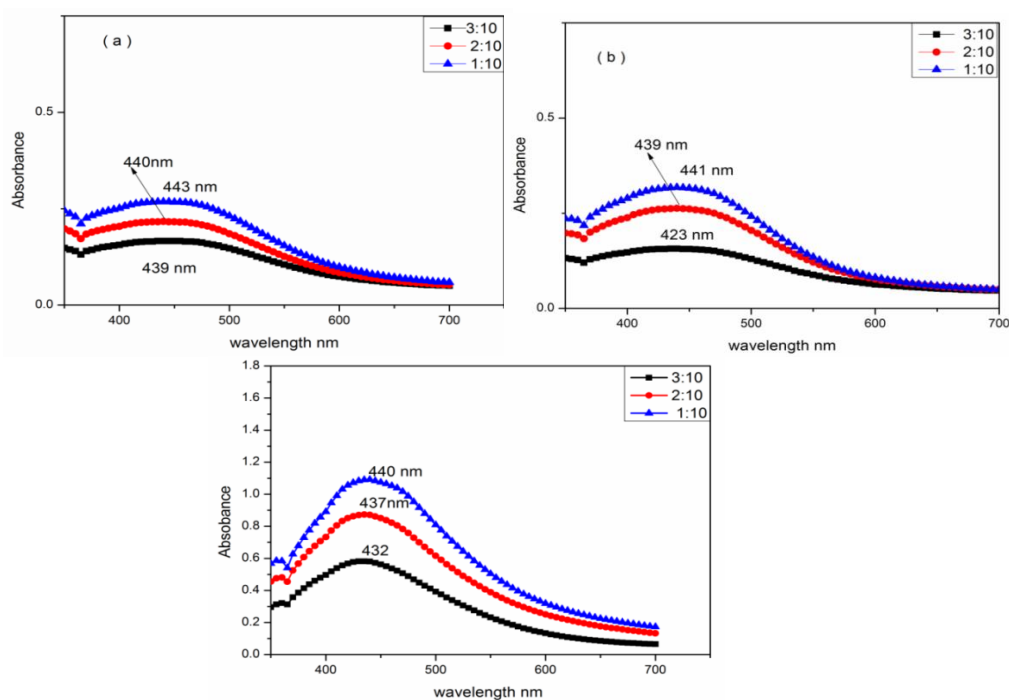


Figure (1): Ultraviolet-visible absorption spectrum of silver nanoparticles prepared using: (a) pepper, (b) Rhamnus, (c) Eucalyptus

3.1.2 X-ray diffraction (XRD)

The X-ray (XRD) patterns of dried silver nanoparticles synthesized using Leaves of *Piper nigrum* , *Ziziphus spina-christi*, and *Eucalyptus globulus* extract showed Bragg reflections representative of the fcc structure of Ag nanoparticles. Figure 2 (a) shows the X-ray diffraction spectra of AgNPS prepared using *Piper nigrum* extract as a function of salt: extract ratio , where a peaks at the angle of 38.2 and 44.4, corresponding to the (111) and (200) planes of silver, respectively after matching with the standard X-ray model of silver (JCPDS No. 04-0783) . The pattern above shows that the diffraction pattern of any peaks cannot be repeated to other materials and this indicates the purity of the prepared sample and free of any additional impurities []. X-ray diffraction measurements showed a different pattern for the samples prepared. The pattern of the 1:10 pattern was better than the other samples. The 3:10 sample did not show exact values compared to the rest of other samples due to the increased concentration of the extract compared to the other samples. The concentration of silver salt, which reduces the proportion of silver particles in the solution, means an increase in the dilution of the resulting solution. Sherrer formula [13] was used to calculate the particle size. It was found that the best size was for model 1:10 which is 20 nm while the ratio of 2:10 was 40nm. Figure 2 (b) shows the X-ray diffraction pattern of nanoparticles where a peak is observed at 2θ equal to 38, and 43 which related to the Silver after its conformity with the standard model of X-ray diffraction of silver (JCPDS No. 04-0783). The X-ray diffraction measurements showed a different pattern of the prepared samples as a function of salt: extract ratio. The diffraction pattern of the prepared at 1:10 better than the other others, while the sample with a ratio of 3: 10 did not show precise values compared to the rest of the samples due to the increased concentration of the extract compared with the concentration of silver salt, which reduces the percentage of silver particles in the solution, which means an increase in dilution of the resulting solution. was used to calculate the granular size where it was found that the best size of the sample prepared at 1:10, while it was 45nm for ratio 2:10. XRD pattern for AgNPs prepared using Eucalyptus was shown in Figure 2 (c). It notice that the higher peak intensity was found for sample prepared using Rhamnus compare with other sample prepared using. The behavior for these sample were found look like to samples prepared using papper and Eucalyptus. Figure 2 (c) illustrated that pattern were same behavior, also was the best compare with other where the intensity was highest for peaks

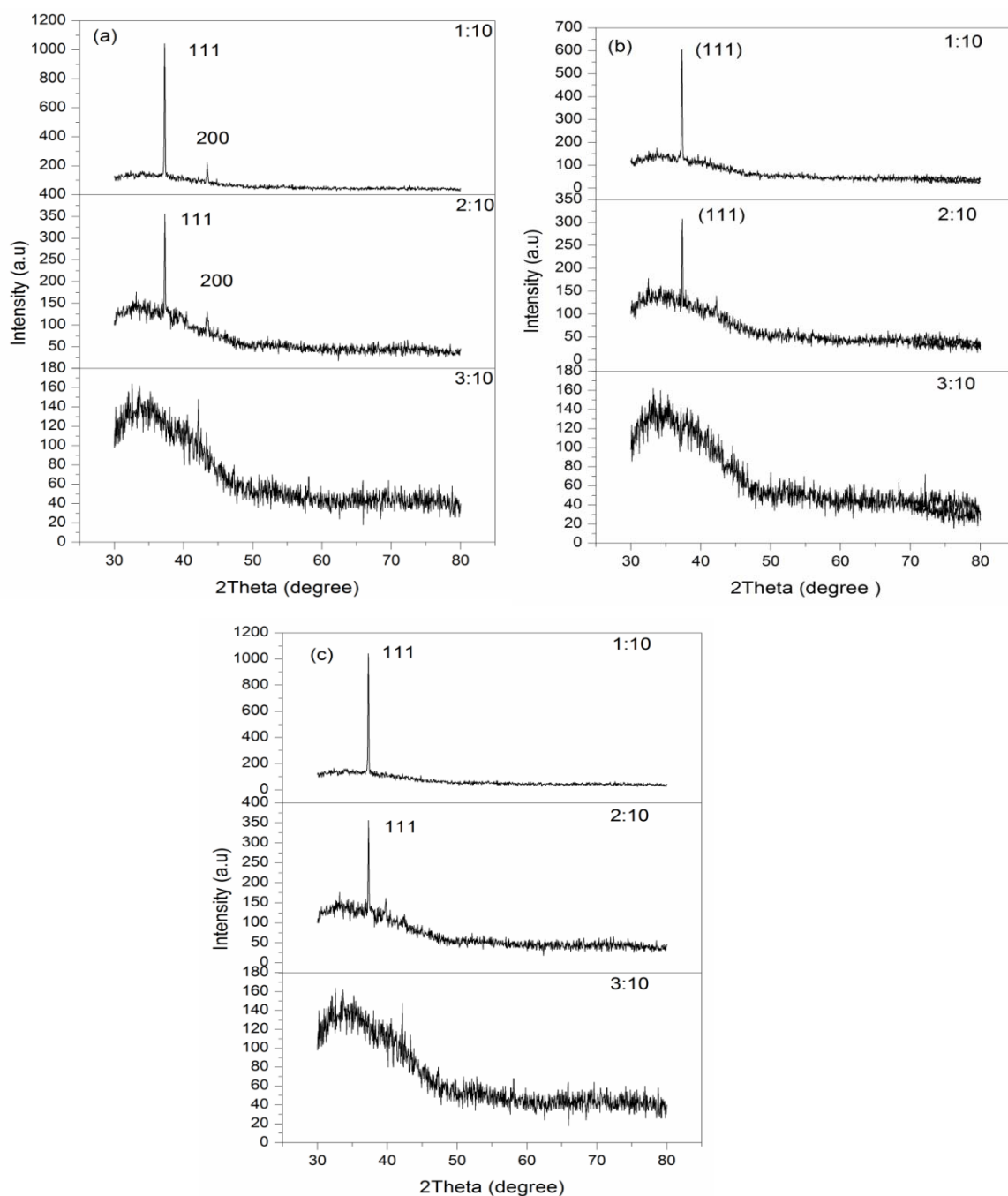


Figure 2: X-ray pattern of silver nanoparticles prepared using: (a) Piper nigrum ,(b) Ziziphus spinachristi, (c) Eucalyptus globulus

4. Field Emission Scanning Electron Microscopy Study

The morphology of green synthesized AgNPs was viewed by FESEM. The FESEM image, as represented in Fig. 4, showed that Particle shapes are usually irregular spheroid and spherical were well dispersed with and particle sizes ranging from 13 to 53 nm. the particle size of nanoparticles through microscopic electron microscopy images, which approximates the values obtained from X-ray diffraction measurements. It is known that the optical and electronic properties of metal nanoparticles

are largely affected by the shape of nanoparticles. This finding strongly confirms that the Rhamnus extract may act as a reduction factor in the production of silver nanoparticles

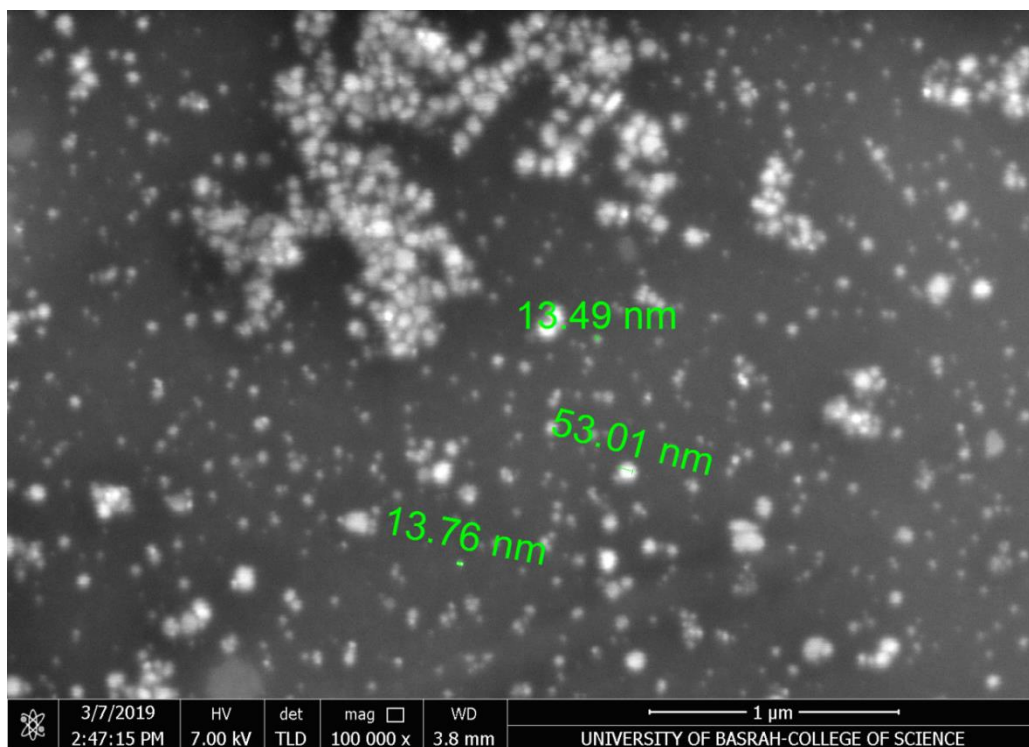


Figure 3: FESEM image of the Ag-NPs

5. Conclusion

Many researchers are highly interested in nanoparticles and the diversity of their fields of application such as electronics, photonics, medicine, and farming. Due to the diversity of organisms ranging from microorganisms to plants, Manufacturing of nanoparticles thus generated has properties that include stable, non-toxic, inexpensive, environmentally friendly and synthesized using green chemistry Unlike conventional chemical and physical processes which often use toxic substances that have the potential to cause environmental toxicity, cellular toxicity, and carcinogenesis. Widely used in the development of nanoparticles, plant use offers a simple, safe and easy to prepare, non-toxic and strong need no preparation or long time to prepare them needed techniques have the potential to commonly use medical procedures for bacteria and fungi techniques. Composite nanoparticles have the potential to be used in the supply of antimicrobial compounds for use as agricultural pesticides In the case of plant extracts, the mechanisms of composition of nanoparticles vary between different plant species.

References

- [1] Laurent, S., Forge, D., Port, M., Roch, A., Robic, C., Vander Elst, L., Muller, R.N., 2010. Magnetic iron oxide nanoparticles: synthesis, stabilization, vectorization, physicochemical characterizations, and biological applications. *Chem. Rev.* 11
- [2] Cheon, J. and G. Horace, 2009. Inorganic nanoparticles for biological sensing, imaging and therapeutics. *J. Mater. Chem.*, 19: 6249-6250.
- [3] Edina C. Wang and Andrew Z. Wang. nanoparticles and their applications in cell and molecular biology. *Integr Biol (Camb)*. 2014 Jan; 6(1): 9–26.
- [4] Chang ALS, Khosravi V, Egbert B. A case of argyria after colloidal silver ingestion. *Journal of Cutaneous Pathology* 2006,(12),3 . ٨٠٩-٨١١

- [5] Chen J, Wang J, Zhang X, Jin Y. Microwave-assisted green synthesis of silver nanoparticles by carboxymethyl cellulose sodium and silver nitrate. *Materials Chemistry and Physics* .2008; 108: 421 – 424.
- [6] Brocchi E.A, Motta M.S, Solorzano I.G, Jena P.K, Moura F.J, *Alternative chemical-based synthesis routes and characterization of nano-scale particles, Mater Sci. Eng. B* 112 (2004) 200–205.
- [7] Janbey R.K., Pati S., Tahir P., Pramanik , *A new chemical route for the synthesis of nano-crystalline α -Al₂O₃ powder, J. Europ. Ceram. Soc.* 21 (2001) 2285–2289.
- [8] E.M. Hunt, K.B. Plantier, M.L. Pantoya, Nano-scale Reactants in the Self-Propagating High-Temperature Synthesis of Nickel Aluminide. *Acta Mater.* 52 (2004) 3183–3191.
- [9] D.G. Yu, Formation of colloidal silver nanoparticles stabilized by Na⁺-poly(γ -glutamic acid)-silver nitrate complex via chemical reduction process. *Colloids Surf. B* 59 (2007) 171–178.
- [10] Y. Tan, Y. Wang, L. Jiang, Thiosalicylic Acid-Functionalized Silver Nanoparticles Synthesized in One-Phase System. *J. Colloid Interface Sci.* 249 (2002) 336–345.
- [11] C. Petit, P. Lixon, M.P. Pileni, In situ synthesis of silver nanocluster in AOT reverse micelles. *J. Phys. Chem.* 97 (1993) 12974–12983.
- [12] S.A. Vorobyova, A.I. Lesnikovich, N.S. Sobal, Preparation of silver nanoparticles by interphase reduction. *Colloids Surf. A* 152 (1999) 375–379.
- [13] Herizchi R1, Abbasi E1, Milani M2,1, Akbarzadeh A. Current methods for synthesis of gold nanoparticles. *Artif Cells Nanomed Biotechnol.* 2016;44(2):596-602.
- [14] Stephen J R and Maenaughton S J, Developments in terrestrial bacterial remediation of metals. *Curr Opin Biotechnol.*, 1999, 10, 230. 5.
- [15] Ernest Merian, Thomas W Clarkson, *Metals and their compounds in the environment : occurrence, analysis, and biological relevance*, Published in 1991 in Weinheim by VCH.
- [16] Nair B and Pradeep T, Coalescence of Nanoclusters and Formation of Submicron Crystallites Assisted by Lactobacillus Strains, *Cryst Growth Des.*, 2002, 2, 293-298.
- [17] Mohanpuria, P.; Rana, N.K.; Yadav, S.K. Biosynthesis of nanoparticles: Technological concepts and future applications. *J. Nanopart. Res.* 2008,10, 507–517.
- [18] Prathap, S.C., Chaudhary, M., Pasricha, R., Ahmad, A., Sastry M, Synthesis of gold nanotriangles and silver nanoparticles using Aloe vera plant extract. *Biotechnol. Prog.*, 22, 577-583 (2006).
- [19] Sontara Konwar Boruah, Prabin Kumar Boruah, Pradyut Sarma, Chitrani Medhi and Okhil Kumar Medhi. Green synthesis of gold nanoparticles using camellia sinensis and kinetics of the reaction. *Advanced Materials Letters.* 2012, 3(6), 481-486.
- [20] Amel Taha, and Mustaffa Shamsuddin. Biosynthesis of Gold Nanoparticles Using Psidium Guajava Leaf Extract. *Malaysian Journal of Fundamental and Applied Sciences.* Vol.9, No.3 (2013) 119-122
- [21] Philip D. Rapid green synthesis of spherical gold nanoparticles using Mangifera indica leaf. *Spectrochim Acta A Mol Biomol Spectrosc* 2010; 77: 807–810.
- [22] Atef A. Hassan, Rasha M. Sayed-Elahl, Noha H. Oraby, Ahmed M.A. El-Hamaky. Metal nanoparticles for management of mycotoxigenic fungi and mycotoxicosis diseases of animals and poultry. 2020, 251-269.
- [23] Chen YH, Tsai CY, Huang PY, et al. Methotrexate conjugated to gold nanoparticles inhibits tumor growth in a syngeneic lung tumor model. *Mol Pharm.* 2007b;4:713–22.
- [24] Tong L, Zhao Y, Huff TB, et al. Gold nanorods mediate tumor cell death by compromising membrane integrity. *Adv Mater.* 2007;19:3136–41.
- [25] Hirsch LR, Stafford RJ, Bankson JA, et al. Nanoshell-mediated near-infrared thermal therapy of tumors under magnetic resonance guidance. *Proc Natl Acad Sci U S A.* 2003b;100:13549–54
- [26] Lesniak A.; Salvati A.; Santos-Martinez M. J.; Radomski M. W.; Dawson K. A.; Aberg C. Nanoparticle Adhesion to the Cell Membrane and Its Effect on Nanoparticle Uptake Efficiency. *J. Am. Chem. Soc.* 2013, 135, 1438–1444

- [27] Zhu M. T.; Li Y. Y.; Shi J.; Feng W. Y.; Nie G. J.; Zhao Y. L. Exosomes as Extrapulmonary Signaling Conveyors for Nanoparticle-Induced Systemic Immune Activation. *Small* 2012, 8, 404–412
- [28] Huo S. D.; Ma H. L.; Huang K. Y.; Liu J.; Wei T.; Jin S. B.; Zhang J. C.; He S. T.; Liang X. J. Superior Penetration and Retention Behavior of 50 nm Gold Nanoparticles in Tumors. *Cancer Res.* 2013, 73, 319–330.
- [29] Dykman L.; Khlebtsov N. Gold Nanoparticles in Biomedical Applications: Recent Advances and Perspectives. *Chem. Soc. Rev.* 2012, 41, 2256–2282
- [30] Mohammad Amzad Hossain, Khulood Ahmed Salim AL-Raqmi, Zawan Hamood AL-Mijizy, Afaf Mohammed Weli, Qasim Al-Riyami. Study of total phenol, flavonoids contents and phytochemical screening of various leaves crude extracts of locally grown *Thymus vulgaris* Asian Pac J Trop Biomed 2013; 3(9): 705-710.
- [31] Marzouk B, Edziri H, Haloui I, Issawi M, Chraief I, El-Ouni M, et al. Chemical composition, antibacterial and antioxidant activities of a new chemo type of Tunisian *Thymus vulgaris* oils growing in Sayada. *J Food Agric Environ* 2009; 7(2): 263-267.
- [32] Arnal-Schnebel B, Hadji-Minaglou F, Peroteau JF, Ribeyre F, De Billerbeck VG. Essential oils in infectious gynaecological disease: a statistical study of 658 cases. *Int J Aromather* 2004;14(4): 192-197.
- [33] Paz Elia, Raya Zach, Sharon Hazan, Sofiya Kolusheva , Ze'ev Porat, Yehuda Zeiri. Green synthesis of gold nanoparticles using plant extracts as reducing agents. *Int J Nanomedicine.* 2014; 9: 4007–4021.
- [34] U. Holzwarth and N. Gibson, The Scherrer equation versus the 'Debye-Scherrer equation. *Nat Nanotechnol* ,28 ,6,534 (2011).
- [35] Gao, S.; Yu, B.; Li, Y.; Dong, W. and Luo, H. Antiproliferative effect of Octreotide on gastric cells mediated by Inhibition of Akt/PKB and telomerase. *World J. Gastroentrol.*,2003, 9: 2362-5.
- [36] Das RK, Sharma P, Nahar P, et al. Synthesis of gold nanoparticles using aqueous extract of *Calotropis procera* latex. *Mat Let.*2011;65:610-3.
- [37] Vijayakumar R, Devi V, Adavallan K, et al. Green synthesis and characterization of gold nanoparticles using extract of anti-tumor potent *Crocus sativus*. *Physica E Low Dimens Syst Nanostruct.* 2011;44:665-71.
- [38] Meena Kumari M, Philip D. Facile one-pot synthesis of gold and silver nanocatalysts using edible coconut oil. *Spectrochim Acta A Mol Biomol Spectrosc.* 2013 Jul;111:154-60
- [39] Mie G.(1908):" Contributions to the optics of turbid media, especially colloidal metal solutions. *Ann Phys* .25:377-445.
- [40] Smitha, SL, Philip, D, Gopchandran, KG: Green synthesis of gold nanoparticles using *Cinnamomum zeylanicum* leaf broth. *Spectrochim. Acta Part A* 74, 735–739 (2009)
- [41] Y. Song, Y.Z. Song, J. Xu. Electrochemical determination of tartaric acid at nano gold/nano carbon modified glassy carbon electrode. *Russ. J. Phys. Chem.A.*86(2012)1458.
- [42] Devi, JS, Bhimba, BV, Ratnam, K: Invitro anticancer activity of silver nanoparticles synthesized using the extract of *Gelidiella* sp. *Int. J. Pharm.Pharm. Sci.* 4, 710–715 (2012).
- [43] Suganya U.S.U., Govindaraju K., Kumar G.G., Prabhu D., Arulvasu C., Dhas S.S., Karthick V., Changmai N. Anti-proliferative effect of biogenic gold nanoparticles against breast cancer cell lines (MDA-MB-231 & MCF-7) *Appl. Surf. Sci.* 2016;371:415–424.
- [44] Wu W, Huang J, Wu L, Sun D, Lin L, Zhou Y, et al. Two-step size-and shape separation of biosynthesized gold nanoparticles. *Sep Purif Technol.* 2013;106 (3):117-22
- [45] Abirami Hariharan, Tajuddin Nargis Begum, Mohamed Hussain Muhammad Ilyas, Hussain Syed Jahangir, Premkumar Kumpat, Shilu Mathew, Archunan Govindaraju, and Ishtiaq Qadri. Synthesis of Plant Mediated gold Nanoparticles using *Azima Tetracantha* Lam. Leaves extract and Evaluation of their Antimicrobial Activities. *Pharmacogn. J.*2016;8(5):507-512.

- [46] Manar E Selim¹, Awatif A Hendi, Gold Nanoparticles Induce Apoptosis in MCF-7 Human Breast Cancer Cells. *Asian Pacific J Cancer Prev*, 13, 1617-1626.
- [47] Jana NR, Gearheart L, Murphy CJ (2001). Wet chemical synthesis of high aspect ratio cylindrical gold nanorods. *J Phys Chem B*, 105, 4065-67 .
- [48] Sun Y, Xia Y (2002). Shape-controlled synthesis of gold and silver nanoparticles. *Science*, 298, 2176-9.
- [49] Nie S, Xing Y, Kim GJ, Simons JW (2007) Nanotechnology applications in cancer. *Annu Rev Biomed Eng* 9: 257-288.
- [50] Chithrani BD, Ghazani AA, Chan WC (2006). Determining the size and shape dependence of gold nanoparticle uptake into mammalian cells. *Nano Lett*, 6, 662-8.
- [51] Chithrani BD, Chan WC (2007). Elucidating the mechanism of cellular uptake and removal of protein coated gold nanoparticles of different sizes and shapes. *Nano Lett*, 7, 1542-50.
- [52] Ali Shakeri-Zadeh, G. Ali Mansoori, A. Reza Hashemian, Hossein Eshghi, Ameneh Sazgarnia, A. Reza Montazerabadi. Cancerous Cells Targeting and Destruction Using Folate Conjugated Gold Nanoparticles. *Dynamic Biochemistry, Process Biotechnology and Molecular Biology*.