ANTIOXIDANT ACTIVITIES FROM LEAVES OF *ELEPHANTOPUS SCABER* L. EXTRACTED USING DIFFERENT SOLVENTS

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ABSTRACT : The present study was carried out to evaluate the antioxidant activities of leaves of *Elephantopus scaber* by using different solvent (ethyl alcohol, acetone and aqueous). The leaves extract from *Elephantopus scaber* using different solvents to extract were evaluated for antioxidant activity by using DPPH method using a different concentrations (1, 10, 20, 30, 40, 60, 80, 100 µg/ml). The results of this method was compared with ascorbic acid as standard. The IC50 value of standard (ascorbic acid) was found to be 20 mg/ml. This study showed the strongest DPPH radical-scavenging activity of ($52.54\pm0.81\%$) inhibition at concentration of 40 mg/ml and IC₅₀ value was 30 mg/ml was found the scavenging activity of ($44.92\pm0.26\%$) for ethyl alcohol extract. Acetone extract of leaves was found at of concentration 60 mg/ml scavenging activity of $53.26\pm0.34\%$. The IC₅₀ value of aqueous extract of leaves was (60μ g/ml) with percentage inhibition of ($54.28\pm0.28\%$) followed by ($44.57\pm0.53\%$) at concentration 40 mg/ml.

Key words : Antioxidant, leaves, Elephantopus scaber L, DPPH, solvents.

INTRODUCTION

Elephantopus scaber L. (*Elephant's foot*) is an important of medicinal plants, which is classified worldwide in all tropical regions (Ho *et al*, 2009). The whole of plant (roots, leaves and flowers) is used medicinally, because of the presence from sesquiterpene lactones, deoxyelephantopin, isodeoxyelephantopin, scabertopin, stigma sterol, epifriedelinol, lupeol and stigma sterol throughout the whole plant (Ho *et al*, 2009).

As the traditional system of medicine, it is reported that the leaves are used as cardiotonic, antipyretic and diuretic activities (Miraldi *et al*, 2001). A decoction of the roots and leaves is useb for diarrhea and emollient and given in dysuria, stomach pain and for dysentery (Duke, 2018).

The aqueous extract of leaves is applied externally to treat eczema and ulcers (Cimanga Kanyanga *et al*, 2018). The waters extract of leaf is applied externally to treat ulcers and eczema. The plant is macerated and used on the surface of wounds to acceleration wound healing activity (Rani *et al*, 2016). It is reported that this species is also used for its hepato protective, antitumor, antiinflammatory activity and wound healing (Asadi-Samani *et al*, 2015).

The growth of this species by seeds is restricted because to poor viability and early death of seedlings in natural environmental conditions (Weber, 2017). In vitroculture is an alternative method of conservation and propagation of this species. Micro propagation of *Elephantopus scaber* L. have been reported through leaves explants (Abraham and Thomas, 2015), but no report have been published regarding plantlet generation from leaves explants, which is the most material for genetic stability of micropropagated plants (Boo *et al*, 2015).

Plants produce a many amount of antioxidants to prevent the oxidative stress caused by reactive oxygen species and are thought to be safer and more healthy than synthetic antioxidants. Biotechnological approaches such as plant tissue culture has been adopted for enhancing bioactive molecules in medicinal plants because to the optimum concentration of growth regulators and nutrients (Franca *et al*, 2016).

This study carried for assess the antioxidant activity of leaves from *Elephantopus scaber* L. extracts using different solvents (ethyl alcohol, acetone and aqueous).

MATERIALS AND METHODS

Collection the plant

The plants collected from Agro Technology Research Station, University Malaysia Perlis Padang Besar, Perlis, Malaysia. The plant has been selected at the flowering stage.

Table 1 : Percentage inhibition of standard (ascorbic acid), leaves extract used solvents (Ethyl alcohol
,acetone and aqueous), at various concentrations (mg/ml) in DPPH scavenging model.

Concentration	Leaves extracts			
	Ethel alcohol	Acetone	Aqueous	Standard
1	28.82±0.38	27.92±0.55	26.87±0.24	33.59±0.19
10	33.36±0.54	30.42±0.42	34.42±0.45	43.89±0.38
20	36.23±0.46	34.04±0.35	38.25±0.48	54.37±0.27
30	44.92±0.26	43.98±0.58	42.55±0.26	61.68±0.38
40	52.54±0.81	45.86±0.28	44.57±0.53	70.04±0.27
60	57.48±0.26	53.26±0.34	54.28±0.28	81.16±0.46
80	60.92±0.52	63.70±0.71	59.34±0.33	88.74±0.19
100	78.73±0.89	70.38±0.28	64.83±0.26	95.62±0.39

Preparation of plant extracts

The selected *Elephantopus scaber* L. was washed thoroughly and separated the leaves from plants then dried under shade. The dried plants material was grounded into fine powder using Waring blender (Cole Parmer, RZ-04245021). The powdered material was extracted by Soxhlet apparatus for 24 hours using different solvents (ethyl alcohol, acetone and aqueous). The solvent was then evaporated using Rotary evaporator at 4°C for further use (Molla, 2015).

Antioxidant activity assay

Determination of antioxidant activity of leaves extract using different solvent were done by DPPH free radical scavenging activity assay.

DPPH free radical scavenging activity assay

Three samples were prepared at the concentration of 1000ig/ml. From the stock solution, different concentration, 1, 10, 20, 30, 40, 50, 60, 80 and 100 µg/ml were prepared and used for antioxidant assay. Ascorbic acid was used as standard for study of DPPH scavenging assay. The stock concentration of 1000 µg/ml was prepared in methanol. From the stock concentration all other concentration ascorbic acid, 1, 10, 20, 30, 40, 50, 60, 80 and 100 µg/ml were prepared.

The antioxidant activity will be measured in terms of hydrogen donating or radical scavenging ability using the stable radical DPPH. Experiments will be carried out according to the method of Blois, with a slight modification. The reduction of the radical is followed by a decrease in the absorbance at 517 nm. A volume of 4 mL of a water or methanolic stock solution of the extracts will put in test tubes and 4 mL of 2 mM DPPH solution will be added. The tubes will be covered use parafilm and keep again at the dark for 1 hours. Absorbance in 517 nm will be measured with a spectrophotometer UVvisu (Jascos V-531) and compared to an vitamin C calibration curve. The results will be express in mg ascorbic acid (vitamin C)/g dried sample. Each assay will carried out in triplicate. The percentage inhibition from the DPPH radical will be calculate by using the formula:

$$I\% = (A0 - A/A0) \times 100$$

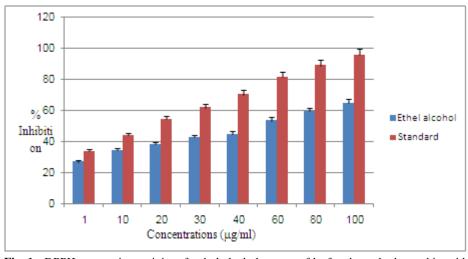
where, I = DPPH inhibition (%), A0 = absorbance of control sample (t = 0 h) and A = absorbance of a tested sample at the end of the reaction (t = 1 h). The amount of sample necessary to decrease the absorbance of DPPH by 50% (IC50) (Gardeli *et al*, 2008; Proestos *et al*, 2013; Gardeli *et al*, 2008; Proestos *et al*, 2013).

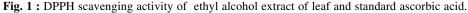
RESULTS AND DISCUSSION

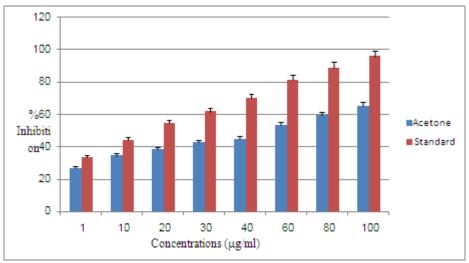
Antioxidant activity assay

In the present study, have evaluated the DPPH free radical scavenging activity of *Elephanthopus scaber* from leaves used a different solvent (Ethyl alcohol, acetone and aqueous). The antioxidant activity of the leaves extracts was evaluated by DPPH radical scavenging protocol. DPPH free radical scavenging activity of leaves extract was compared with ascorbic acidas standard (Table 1).

Ethel alcohol extract of leaf part of *Elephanthopus* scaber showed the highest DPPH scavenging activities at concentration of (100µg/ml) where the scavenging activity was (78.73±0.89%) followed by (70.38±0.28%) of acetone extract then (64.83±0.26%) for aqueous extract while at the same concentration, scavenging activity of ascorbic acid was (95.62 \pm 0.39%). The IC₅₀ value of standard was (20µg/ml) with percentage inhibition of (54.37±0.27%). For Ethel extract, Acetone and Aqueous extract the IC₅₀ value was (40µg/ml) with inhibition percentage of (52.54±0.81%), (45.86±0.28%) and (44.57 \pm 0.53%), respectively. The IC₅₀ value of ethyl alcohol, acetone and aqueous extracts was 80µg/ml with inhibition percentage of $(60.92\pm0.52\%)$, $(63.70\pm0.71\%)$, (59.34±0.33%), respectively. The inhibitory effect of the leaves extracts on DPPH radicals followed dose-dependent manner (Figs. 1, 2 and 3).







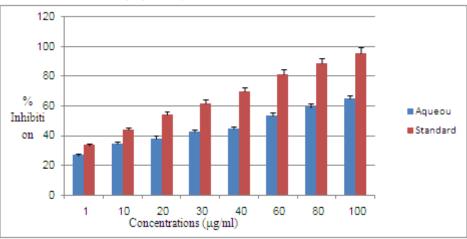


Fig. 2 : DPPH scavenging activity of acetone extract of leaf and standard ascorbic acid.

Fig. 3 : DPPH scavenging activity of aqueous extract of leaf and standard ascorbic acid.

REFERENCES

Abraham J and Thomas T D (2015) Plant regeneration from organogenic callus and assessment of clonal fidelity in *Elephantopus scaber* Linn., an ethnomedicinal herb. *Physiology and Molecular Biology of Plants* **21**(2), 269-277.

Asadi-Samani M, Kafash-Farkhad N, Azimi N, Fasihi A, Alinia-

Ahandani E and Rafieian-Kopaei M (2015) Medicinal plants with hepatoprotective activity in Iranian folk medicine. *Asian Pacific Journal of Tropical Biomedicine* **5**(2), 146-157.

Boo K H, Cao D V, Pamplona R S, Lee D, Riu K-Z and Lee D-S (2015) In vitro plant regeneration of Aster scaber via somatic embryogenesis. *Bioscience, Biotechnology and Biochemistry* 79(5), 725-731. Cimanga Kanyanga R, Malika Bool-Miting F, Tona Lutete G, Kambu Kabangu O, Vlietinck A and Pieters L (2018) Antibacterial screening of aqueous extracts of some medicinal plant and their fractions used as antidiarrheal agents in Kinshasa-Democratic Republic of Congo. *World Journal of Pharmacy and Pharmaceutical Sciences* 7(1), 223-242.

Duke J A (2018) Amazonian ethnobotanical dictionary: CRC press.

- Franca J V, Queiroz M S R, do Amaral B P, Simas N K, da Silva N C B and Leal I C R (2016) Distinct growth and extractive methods of Acmella oleracea (L.) RK Jansen rising different concentrations of spilanthol: An important bioactive compound in human dietary. Food Research International 89, 781-789.
- Gardeli C, Vassiliki P, Athanasios M, Kibouris T and Komaitis M (2008) Essential oil composition of *Pistacia lentiscus* L. and *Myrtus communis* L.: Evaluation of antioxidant capacity of methanolic extracts. *Food Chemistry* **107**(3), 1120-1130.

Ho W Y, Ky H, Yeap S K, Rahim R A, Omar A R, Ho C L and

Alitheen N B (2009) Traditional practice, bioactivities and commercialization potential of *Elephantopus scaber* Linn.

- Miraldi E, Ferri S and Mostaghimi V (2001) Botanical drugs and preparations in the traditional medicine of West Azerbaijan (Iran). *Journal of Ethnopharmacology* **75**(2-3), 77-87.
- Molla Y (2015) Evaluation of the antibacterial activity of the solvent fractions of the leaves of *Rhamnus prinoides* L'Herit (Rhamnaceae). Addis Ababa University Addis Ababa, Ethiopia.
- Proestos C, Lytoudi K, Mavromelanidou O K, Zoumpoulakis P and Sinanoglou V J (2013) Antioxidant capacity of selected plant extracts and their essential oils. *Antioxidants* **2**(1), 11-22.
- Rani S, Amanjot H, Singh N, Gautam S P and Kaur S (2016) Formulation, optimization and evaluation of dendricream for wound healing activity of *Artemisia indica*.
- Weber E (2017) Invasive plant species of the world: a reference guide to environmental weeds: CABI.